

A COMPARISON OF RNA HOMOLOGY-DETECTING SOFTWARE

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Bioinformatics Masters Thesis
December 2008

The problem of RNA secondary structure prediction

- Primary structure does not necessarily imply secondary structure
- Secondary structure better conserved than primary sequence for RNA
- Common secondary structures can show that two RNAs are related, where sequence alignment failed

Covariance Models are one approach

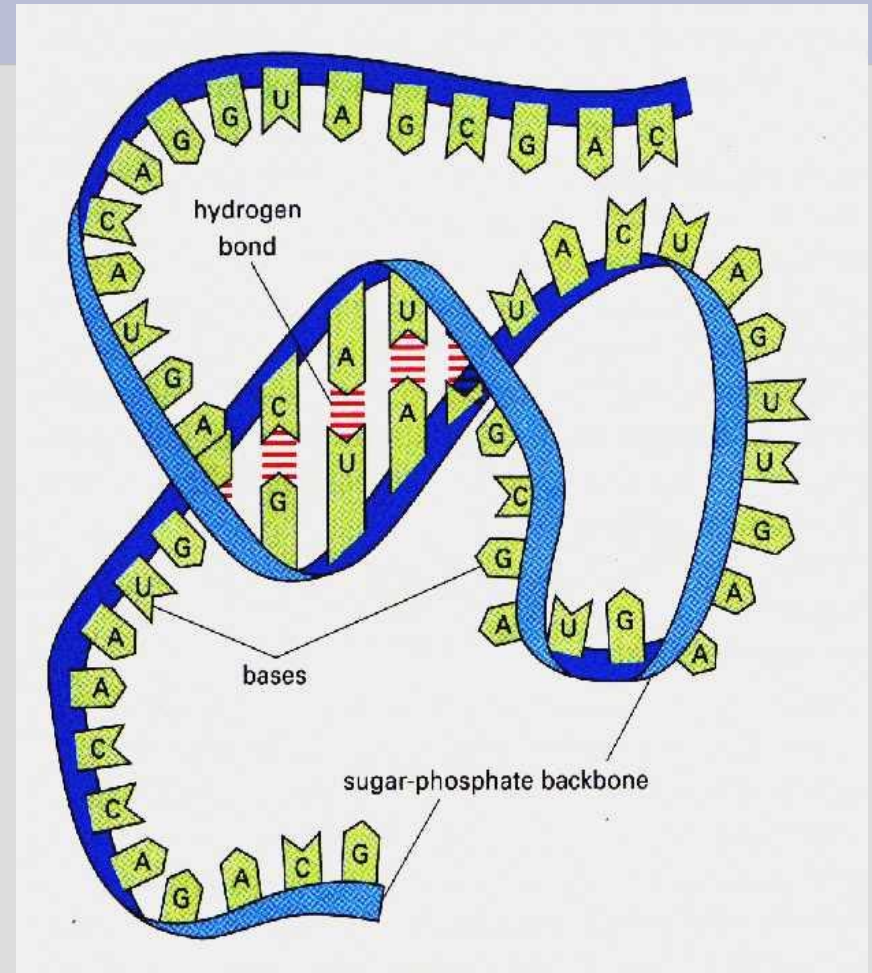
- Probabilistic model
- Describes secondary structure and primary sequence
- Can be used for secondary structure prediction, multiple sequence alignment, database similarity searching
- Intended to find RNAs where sequence alignments alone would not work as well

Application of Covariance Models to RNA

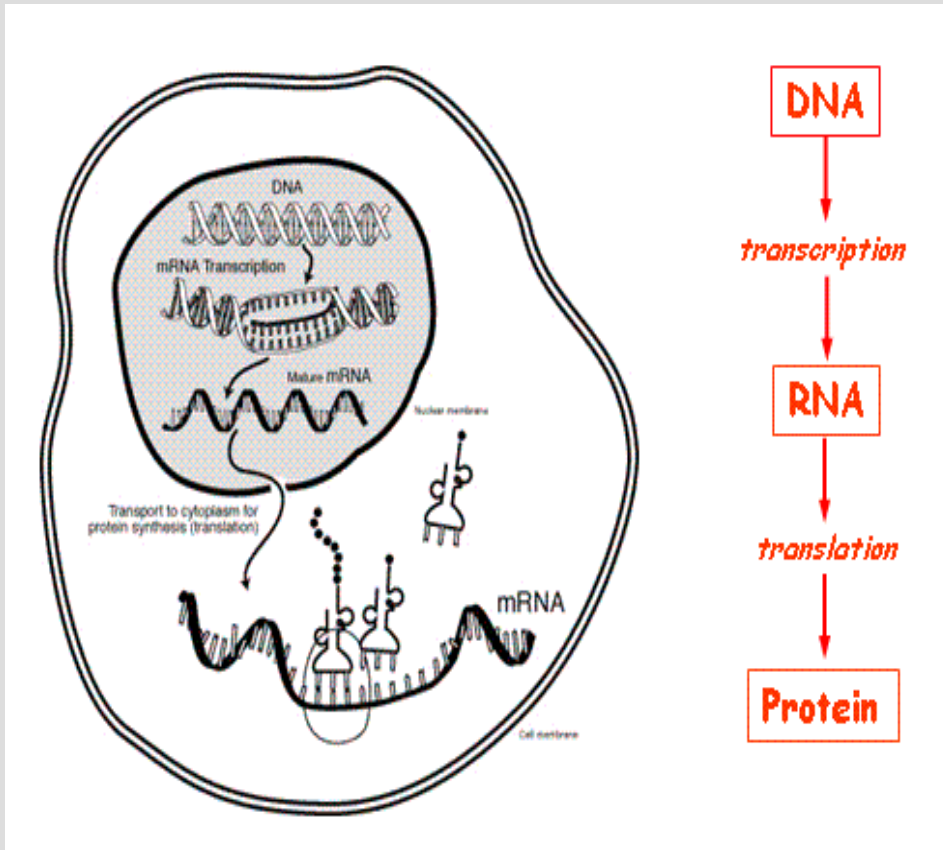
- I. Background of topic, both from biology and computer science perspective
- II. Survey of software using CMs
- III. Databases used
- IV. Methods & results

RNA background

- RNA: Once thought to be mere messenger molecule, but now known to be both an information carrier and an enzymatically active molecule
- Some have suggested it is the original biological molecule (the “RNA world” theory)



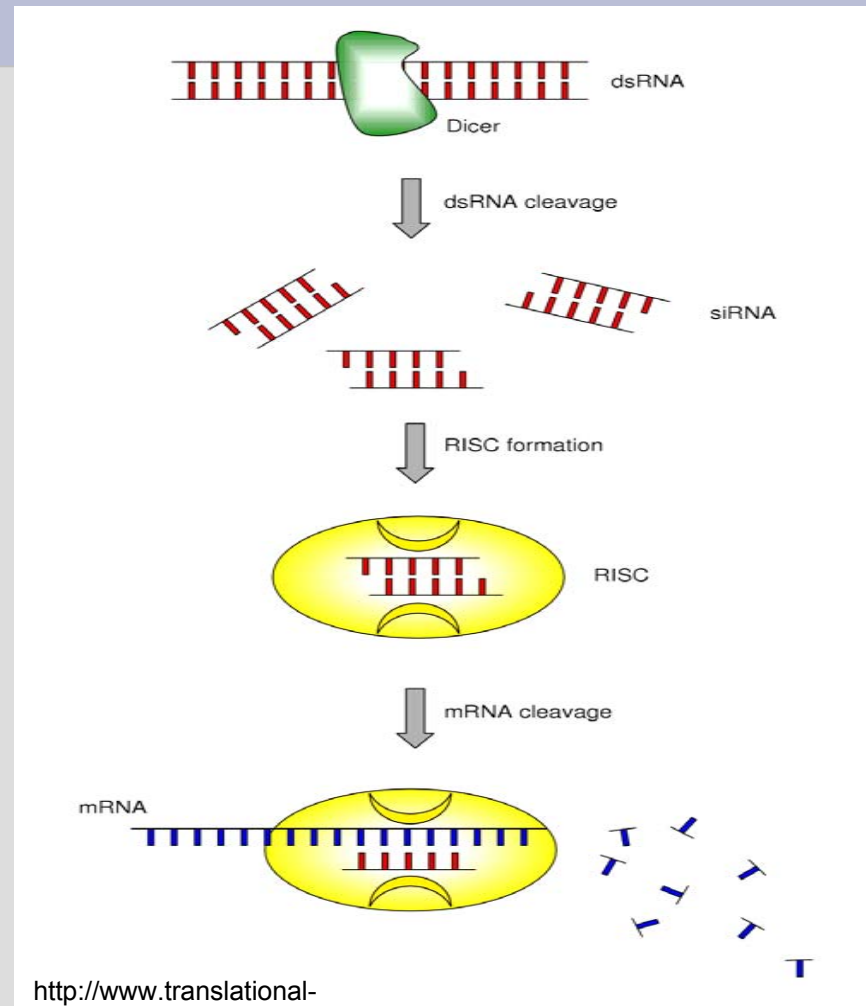
The Central Dogma



- DNA—main information carrying molecule, copies itself in the process of replication
- DNA is “copied” onto mRNA, the process of transcription
- RNA is then used to create protein, the process of translation
- No information flows from protein to DNA
- Translation assisted by tRNA and rRNA

But the Central Dogma might be a little too simple ...

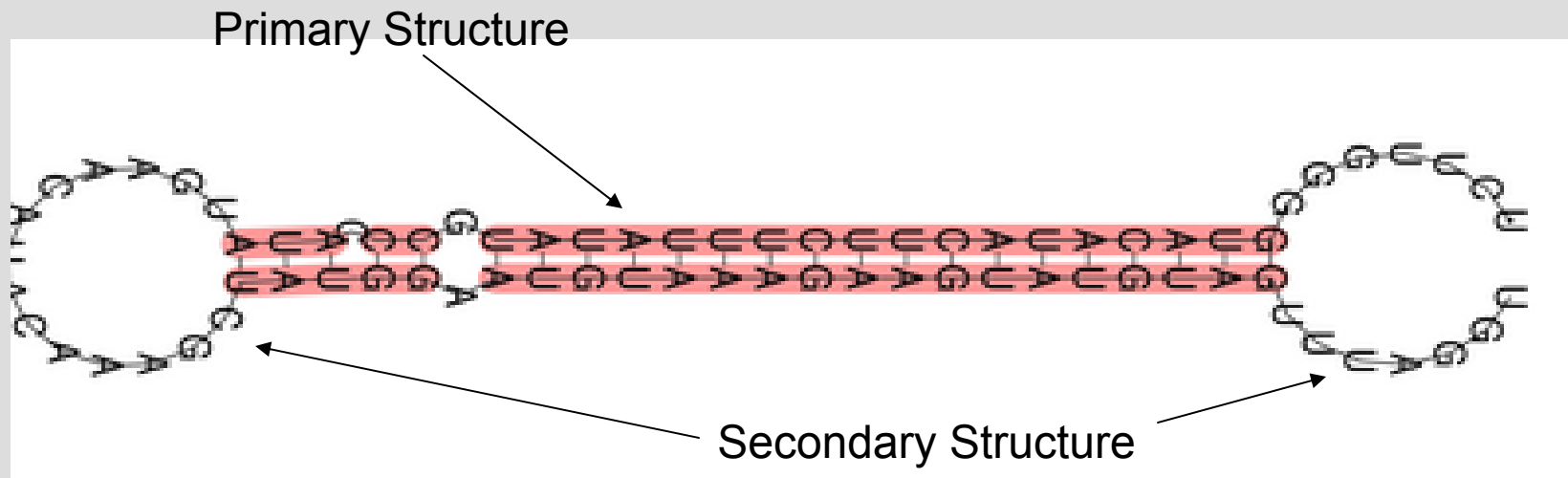
- RNA has a much more active role in all facets of cell life than previously realized, in regulation, gene expression, etc.
- Epigenetics: heritable traits that do not involve a change in DNA sequence
- Some epigenetic heredity is due to RNA interference, by methylating certain DNA sites, or activating/degrading certain RNAs and proteins



<http://www.translational-medicine.com/content/2/1/39/figure/F1?highres=y>

RNA structure background

- Primary Structure
- Refers to sequence of nucleic acid residues
- Secondary Structure
- Refers to a number of small subunits RNA tends to fold into (like stem-loops)



The miRNA *mir-1*.

<http://rfam.sanger.ac.uk/family?acc=RF00103>

Typical RNA secondary structures

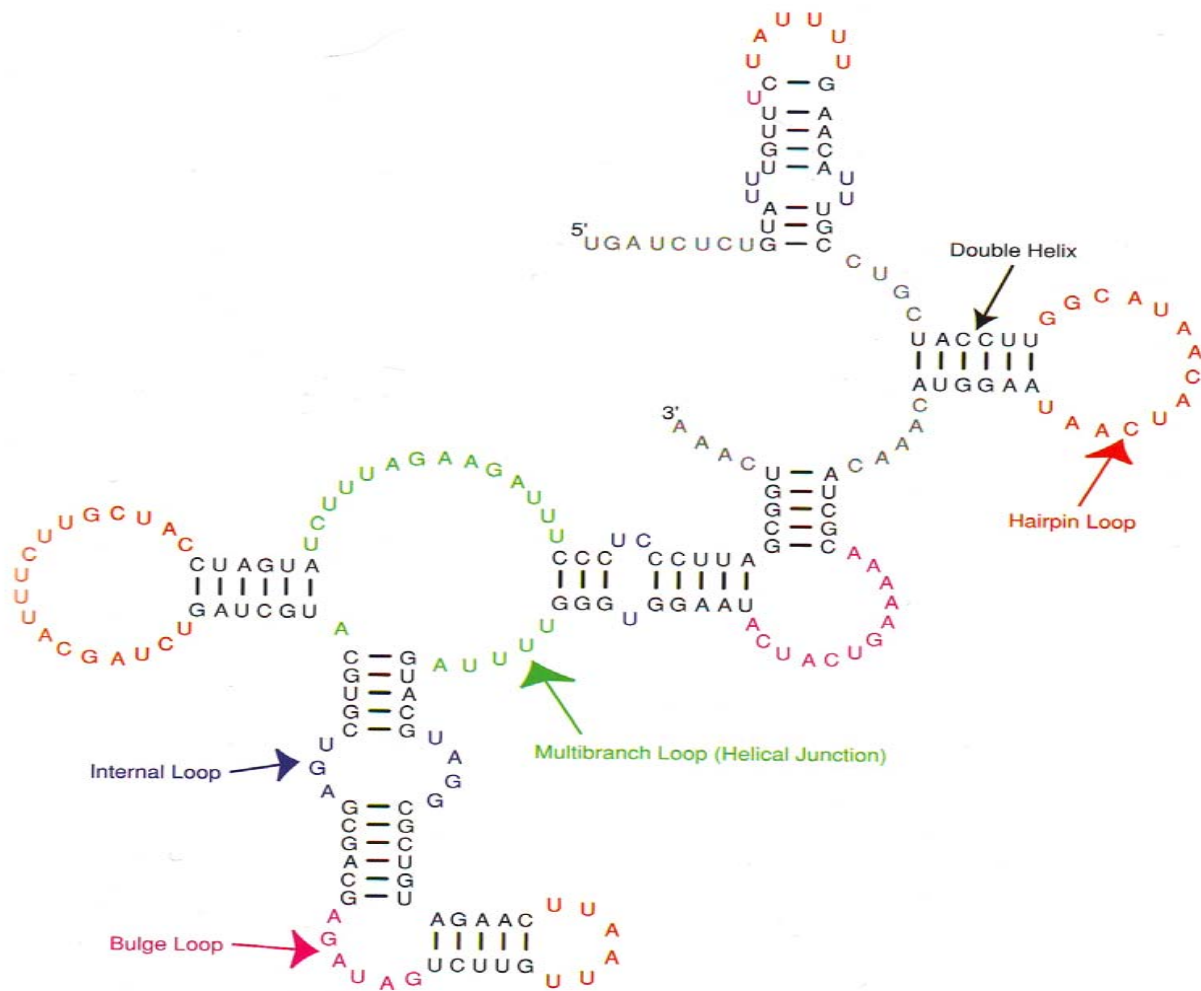


FIGURE 6.2 The RNA secondary structure of the 3' UTR from the *D. sucinea* R2 element (Lathe & Eickbush, 1997; Mathews et al., 1997). Base pairs in nonhelical regions, known as loops, are colored by type of loop.

From Baxevanis' Bioinformatics, 3rd edition.

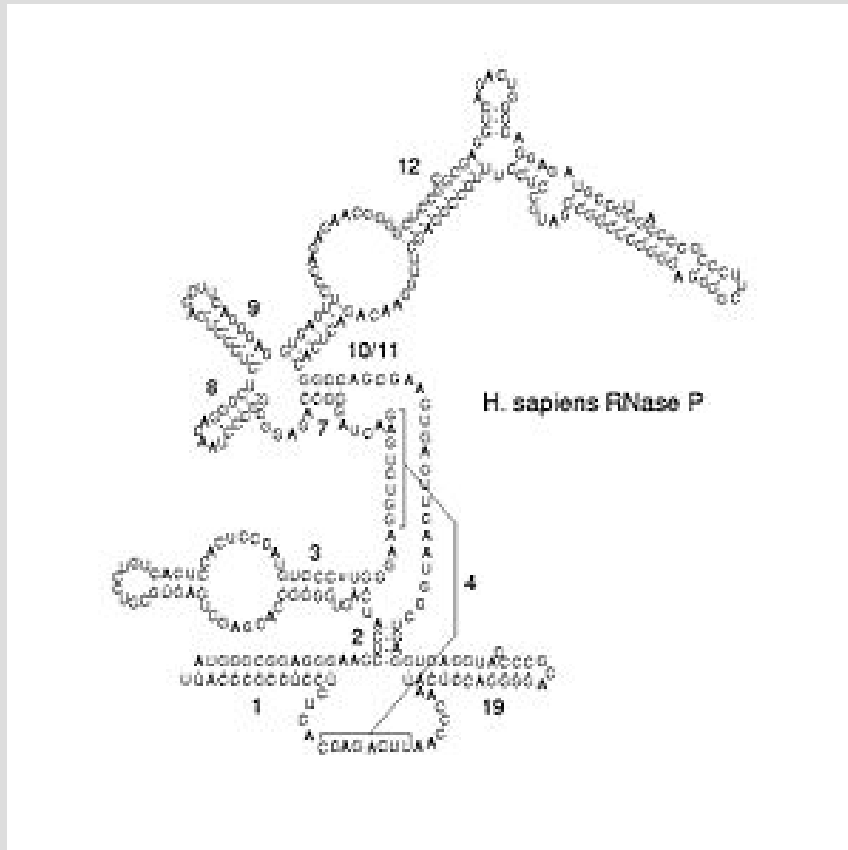
Types of RNA

- Messenger RNA
- Transfer RNA
- Ribosomal RNA
- Non-coding RNAs
 - Ribozymes & riboswitches
 - Cis-regulatory elements
 - Micro RNAs
 - siRNAs and shRNAs
 - snRNAs and snoRNAs
 - Telomerase RNA

ncRNAs

- ncRNAs: Functional, not information carriers
- In essence, all RNA that isn't messenger
- Other than the well known transfer and ribosomal RNAs, it was once dismissed as “junk”
- Now known to have critical regulatory functions (as cis-regulatory elements or gene expression regulating miRNAs)

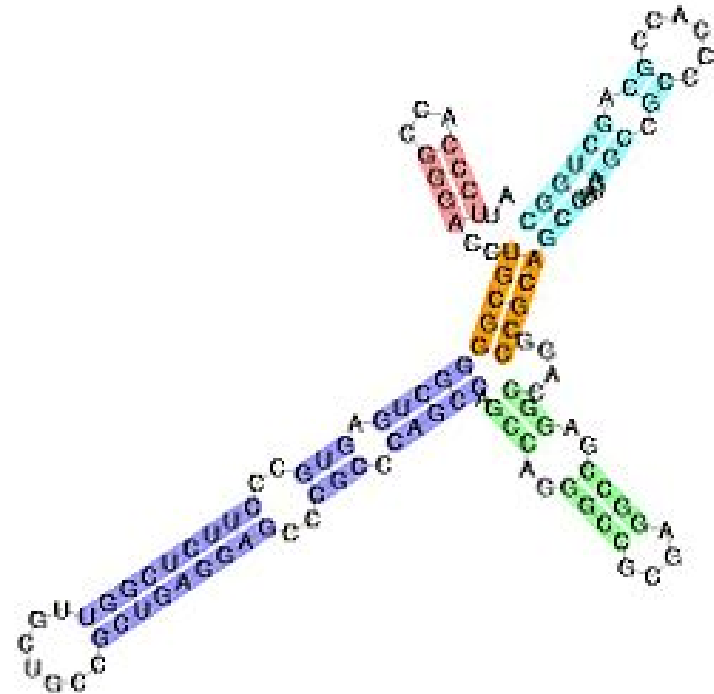
Ribozymes & Riboswitches



- Ribozymes: RNAs that function like enzymes (rnase P, self-cleaving RNA, possibly ribosomes)
- Riboswitches: untranslated segments attached to mRNA that let an mRNA self-regulate itself; common in bacteria

Cis-regulatory elements

- Cis regulation: gene produces functional RNA that regulates genes on the same strand of DNA (as opposed to trans regulation, which acts on distant strands)
- Attach to binding site and influence transcription
- Sequences in the tens to hundreds
- Others influence RNA replication

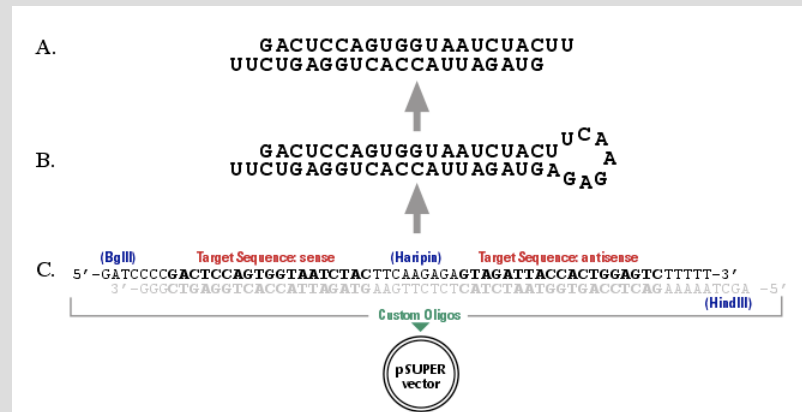


Apolipoprotein B (apoB) 5' UTR cis-regulatory element

<http://rfam.sanger.ac.uk/family?acc=RF00463>

siRNAs & shRNAs

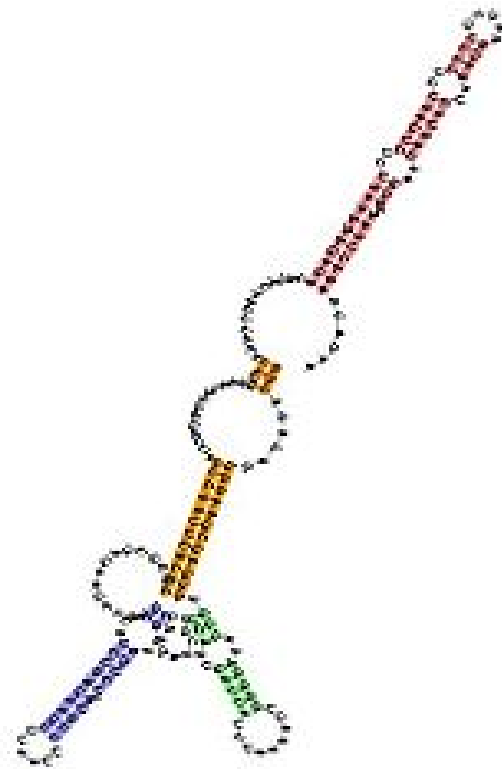
- Small interfering RNAs: function in RNA interference
- Formed from precursors, small hairpin RNAs
- Industry appears to be very interested in these



<http://www.oligoengine.com/products/pSUPER.html>

snRNA and snoRNA

- snRNA: small nuclear RNA
- Active as regulators, splicing agents, telomere maintenance
- Major snRNA class are snoRNAs, small nucleolar RNAs
- Aid in the nucleolus' main function: ribosome creation
- Form RNA-protein complexes (snoRNPs)
- Act via methylation and pseudouridylation (the isomerisation of uridine)



The snoRNA U3.

<http://rfam.sanger.ac.uk/family?acc=RF00012>

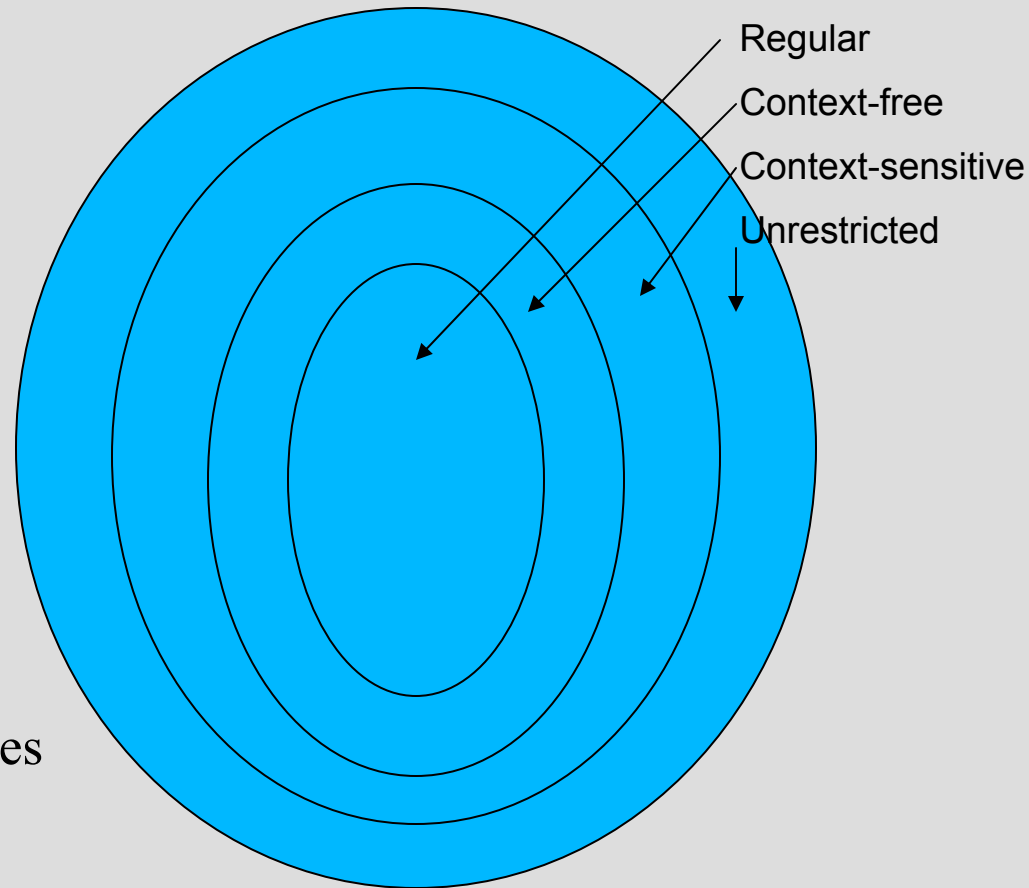
Covariance Models

- Algorithm: Sequence of instructions that must be performed to solve a well-formulated problem (how computer programs accomplish their work)
- Dynamic programming: type of algorithm that breaks problems into smaller problems (can lead to huge complexity)
- DP is used quite a bit to solve RNA secondary structure prediction problems

Covariance Models

Background: Grammars

- Grammar: In computer science terms, a set that describes the possible words or statements in a language
- Chomsky hierarchy of grammars:
 - Regular
 - Context-free
 - Context-sensitive
 - Unrestricted (phase structure)
- Automata: An abstract computational device that describes individual grammars



Covariance Models

Background: Grammars

- Regular grammars: Generate sequence from left to right, and are thus useful for modeling primary sequence
- Context-free grammars: Originally devised to describe natural languages, they have rules that allow the grammars to make correlations between ends of sentences—useful for RNA, where sequence differences may not imply secondary structure differences

Covariance Models

Background: Grammars

- Context-sensitive grammars: Grammars that have additional rules involving nonterminal character replacements that differentiate them from context-free
- Stochastic grammars: Probabilistic grammars where characters are given scores based on consensus of how a grammar is thought to work; every Chomsky hierarchy grammar can have a stochastic form

Covariance Models Background: Stochastic Grammars

- Useful for biological analysis, since there are numerous grammatical exceptions in DNA/RNA; a probabilistic model can account for exceptions
- Example: sequence profiles that contain enough specificity to find distantly related family member
- Hidden Markov Model profiles are a widely used type of stochastic grammar

Covariance Models Background: Stochastic Grammars & CMs

- Covariance models are another type of stochastic grammar based profile
- Unlike HMMs, they can be used to predict secondary structure
- Are the “SCFG analogue of profile HMMs”
- Specify a repetitive tree-like SCFG architecture
- Detailed, complex probabilistic models

Software & Databases Used

- CM-using software:
 - The Infernal suite
 - CMfinder
- CM-using software:
 - CARNAC
 - miRNAMiner
 - BLAT
- Databases used:
 - miRBase
 - Rfam
 - RNA strand
 - UCSC Genome Browser
 - ENSEMBL
 - NCBI Genome

The Infernal suite

- Cmalign
- Cmbuild
- Cmcalibrate
- Cmemit
- Cmscore
- Cmsearch
- cmstat



Infernal: inference of RNA alignments

[infernal home](#) | [rfam database](#) | [eddy lab](#) | [janelia farm](#)

Overview

Infernal ("INFERENCE of RNA Alignment") is for searching DNA sequence databases for RNA structure and sequence similarities. It is an implementation of a special case of profile stochastic context-free grammars called *covariance models* (CMs). A CM is like a sequence profile, but it scores a combination of sequence consensus and RNA secondary structure consensus, so in many cases, it is more capable of identifying RNA homologs that conserve their secondary structure more than their primary sequence.

The latest release of Infernal is [1.0rc3 \[24 Sept 2008\]](#). This is the third release candidate for Infernal 1.0.

Documentation:

- [User's Guide \[PDF, 106 pages\]](#).
- [OOREADME](#) from the current release.

Download:

<http://infernal.janelia.org/>

```
Fred@Fred_HP_Dorf ~/infernal-1.0rc3
$ cd lab
Fred@Fred_HP_Dorf ~/infernal-1.0rc3/lab
$ cmsearch -g let_7_full.cm cow2.fa
# cmsearch :: search a sequence database with an RNA CM
# INFERNAL 1.0rc3 (September 2008)
# Copyright (C) 2008 HHMI Janelia Farm Research Campus
# Freely distributed under the GNU General Public License (GPLv3)
# -----
# command: cmsearch -g let_7_full.cm cow2.fa
# date: Sun Nov 23 02:09:59 2008
# num seqs: 1
# dbsize(Mb): 0.004140
# Pre-search info for CM 1: let_7_full.sto-1
# -----
#                               cutoffs                predictions
# -----
# rnd  mod  alg  cfg  beta  E value  bit sc  surv  run time
# ---  ---  ---  ---  ---  ---
# 1    cm  ins  glc  1e-15  1.000    1.59    0.0196  00:00:07.19
# -----
CM: let_7_full.sto-1
>15
```

The Infernal homepage

Infernal in Cygwin

CMfinder



University of Washington
Computer Science & Engineering

News: CMfinder software [download](#) is available now.

You need help adjusting the parameters of CMfinder? Please read the [manual](#). Questions? [send mail](#)

You can run CMfinder using 2 sets of parameters at one time

First configuration:

Number of stem-loops

Number of motifs <10

Minimum length of motif >15

Maximum length of motif <150

Number of Candidates <100

Expected fraction of sequences containing the motif 0~1

Second configuration:

Number of stem-loops

Number of motifs < 10

Minimum length of motif > 15

Maximum length of motif < 150

Number of Candidates < 100

Expected fraction of sequences containing the motif 0~1

Paste Sequences (*) in FastA Format

Please limit your dataset to
4~60 sequences with length < 500bp

Or Upload a FastA file (*):

Post process options: Merge motifs Remove Redundant motifs

miRNAm iner

MIRNAMINER

miRNAm iner is a web-based tool used for homologous miRNA gene search in several species. Given a search query, candidate homologs are identified using BLAST search and then tested for their known miRNA properties, such as secondary structure, energy, alignment and conservation, in order to assess their fidelity. Default parameters are stringent, though these can be relaxed. miRNAm iner can be used prior to depositing novel miRNAs in public databases (such as miRbase).

NEW: See also [miRviewer](#), a global view of homologous miRNA genes in many species.

[Updates](#) | [Discovered miRNAs](#) | [Help](#)

max RNA fold energy (delta G)

minimal length of precursor sequence (nt)

maximal length of precursor sequence (nt)

number of results to report

Require seed conservation in mature miRNA (nt 2-8)

Target genomes (hold Ctrl and click to select more than one):

- Human (homo sapiens)
- Mouse (mus musculus)
- Rat (rattus norvegicus)
- Chimp (pan troglodytes)
- Dog (canis familiaris)

MicroRNA precursor sequence (required):

Mature microRNA sequence (required):

Your email address-miRNAm iner will send results to that email (optional)

When you're ready, press **Submit** and wait for the result screen (search time depends on the number of requested results and the number of searched genomes)

<http://groups.csail.mit.edu/pag/mirnaminer/>

CARNAC

carnac :: RNA structure inference

[home](#) | [web server](#) | [help](#) | [examples](#) | [retrieve result with an ID](#)

Enter a **name** for the sequences (*optional*):

Paste your RNA sequences in FASTA format [\[?\]](#)

or

upload a file

- eliminate redundant sequences [\[?\]](#)
- take GC content into account [\[?\]](#)
- allow isolated stems (may be slow) [\[?\]](#)

Enter your **E-mail** address (*optional*):

<http://bioinfo.lifl.fr/RNA/carnac/carnac.php>

BLAT

new **SETUP** CONFIG RESULTS DISPLAY

refresh **Online Help**

Important Notice

We now used Blat as our default DNA search. This will make your query faster.

Enter the Query Sequence

Either Paste sequences (max 30 sequences) in FASTA or plain text:

Or Upload a file containing one or more FASTA sequences

 Browse...

Or Enter a sequence ID or accession (EMBL, UniProt, RefSeq)

 Retrieve

Or Enter an existing ticket ID:

 Retrieve

- dna queries
- peptide queries

Summary

▶ setup

⌚ Not yet initialised

▶ configure

⌚ Not yet initialised

▶ results

⌚ Not yet initialised

▶ display

⌚ Not yet initialised

Select the databases to search against

Select species:

Use 'ctrl' key to select multiple species

dna database

peptide database

Select the Search Tool

configure ▶ **RUN ▶**

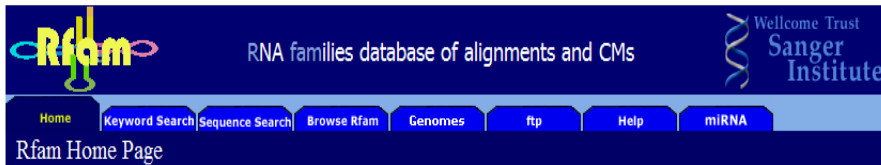
Search sensitivity:

Optimise search parameters to find the following alignments

<http://www.ensembl.org/Multi/blastview>

Rfam

Rfam 8.0



Switch to the new Rfam website

We are currently developing a [new website](#) for Rfam. All new data releases will be available through the new site, but this website will remain on Rfam release 8.1 and will no longer be updated.

If you would like to see data from Rfam release 9.0, please use the new site, at <http://rfam.sanger.ac.uk/>

Rfam is a joint project involving researchers based at the [Wellcome Trust Sanger Institute](#), Cambridge, UK and [Janelia Farm](#), Ashburn, VA, USA. Rfam is a large collection of multiple sequence alignments and covariance models covering many common non-coding RNA families. For each family in Rfam you can:

<http://www.sanger.ac.uk/Software/Rfam/>

Version 8.1

October 2007, 607 families

Enter your keyword(s) here

Enter an EMBL name or accession number

Rfam 9.0

Rfam 9.0 :: Home

The Rfam database of RNA alignments and CMs



[rfam consortium \(cambridge\)](#) | [rfam mirror \(janelia\)](#) | [infernai](#) | [eddy lab](#) | [janelia farm](#)
[home](#) | [browse rfam](#) | [sequence search](#) | [help](#)

Rfam 9.0 (July 2008, 603 families)

Rfam is a collection of multiple sequence alignments and covariance models covering many common non-coding RNA families. The main use of Rfam is as a source of RNA multiple alignments with consensus secondary structure annotation in a consistent format. In conjunction with the **Infernal** software package, Rfam covariance models (CMs) can be used to search genomes or other DNA sequence databases for homologs to known structural RNA families.

Rfam makes use of a large amount of available RNA alignment data, especially published multiple sequence alignments, and repackages these data in a single searchable and sustainable resource. We have made every effort to credit individual sources on each family page (see the [help page](#) for a list). If you find any of the data presented here useful, please also be sure to credit the primary source.

Rfam is produced by the Rfam Consortium, a collaboration between researchers at the **Wellcome Trust Sanger Institute** near Cambridge, UK, the **University of Manchester** in Manchester, UK, and **HHMI Janelia Farm** near Washington, DC.

BROWSE RFAM View Rfam annotation and alignments.

<http://rfam.janelia.org/>

miRBase



miRBase::Sequences

MANCHESTER
1824



[Home](#) [Search](#) [Browse](#) [Genomics](#) [Help](#) [Download](#) [Submit](#) [miRBase](#)

[Search](#)

Search miRBase::Sequences

By miRNA identifier or keyword

Enter a miRNA accession, name or keyword:

[Submit Query](#)

[Reset](#)

[Example](#)

By genomic location

Select organism, chromosome and start and end coordinates. Leave the start/end boxes blank to retrieve all miRNAs on the selected chromosome.

Choose species:

Chr:

Start:

End:

[Get sequences](#)

For clusters

Select organism and the desired inter-miRNA distance.

Choose species:

Inter-miRNA distance:

10000

[Get clusters](#)

By sequence

RNA Strand

RNA STRAND v2.0 - The RNA secondary STRucture and statistical ANalysis Database

[[Home](#) | [Search](#) | [Analyse](#) | [Submit structures](#) | [News](#) | [Help](#)]

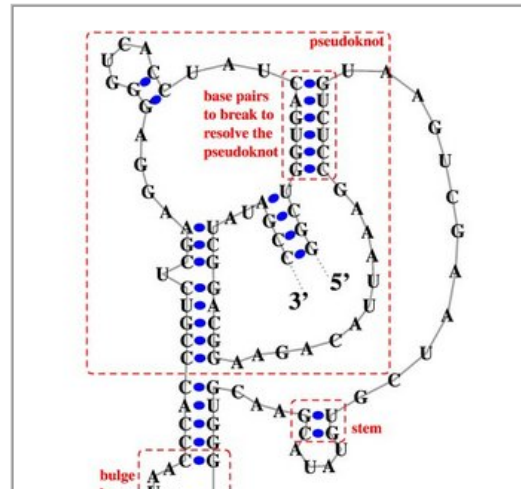
RNA STRAND contains *known RNA secondary structures* of any type and organism. The ultimate goal of this database is to incorporate a comprehensive collection of known RNA secondary structures, and to provide the scientific community with simple yet powerful ways of **analysing, searching and updating** the proposed database.

Current holdings: [4666](#) secondary structures in total.

Search	Search for RNA STRAND entries, supports multiple search criteria
Analyse	Analyse one or a group of RNA secondary structures
Submit	Submit new RNA secondary structures to RNA STRAND

updates on new the database
 tations of RNA STRAND
 utput fields, also
 via the '?' links on any
 ND page

STRAND structures
 nd link to source
 tein Data Bank
 CRW Site



Structural feature occurrences in RNA STRAND		
#RNAs	#Occurrences	Structural motif
2333	6746	Pseudoknots
3582	17537	Multibranched loops
2992	35650	Internal loops
2898	31392	Bulge loops
4575	43442	Hairpin loops
2296	48730	Non-canonical base pairs

Most common RNA types in RNA STRAND	
# RNAs	RNA type
726	Transfer Messenger RNA
723	16S Ribosomal RNA
707	Transfer RNA
470	Ribonuclease P RNA
450	Synthetic RNA
394	Signal Recognition Particle RNA

Provenance of RNA STRAND structures	
#RNAs	Source and link to source
1059	RCSB Protein Data Bank
1056	Gutell Lab CRW Site
726	tmRNA Database
622	Sprinzl tRNA Database
454	RNase P Database
383	SRP Database
313	Rfam Database
53	Nucleic Acid Database

RmotifDB

RmotifDB

A Database of RNA Structural Motifs



Home

Software

BlockMatch

Help

Contact

Search by block (by entering a query block or motif in Stockholm format)

OR Upload from file :

Display top hits

E-mail: (optional)

Data Collection: microRNAs

Family: let-7 (RF00027)

Summary

let-7 microRNA precursor [Edit Wikipedia Entry](#)

The Rfam group coordinates the annotation of Rfam families in [Wikipedia](#). You [here](#). [More...](#)

The **Let-7 microRNA precursor** was identified from a study of developmental timing in *C. elegans*,^[1] and was later shown to be part of a much larger class of **non-coding RNAs** termed **microRNAs**.^[2] miR-98 microRNA precursor from human is a let-7 family member. Let-7 miRNAs have now been predicted or experimentally confirmed in a wide range of species (MIPF000002). miRNAs are transcribed as pri-miRNAs, which are processed in the nucleus by *Drosha* and *Pasha* to hairpin structures of about ~70 nucleotide called pre-miRNAs. These precursors are exported to the cytoplasm by *exportin5*, where they are subsequently processed by the enzyme *Dicer* to a ~22 nucleotide mature miRNA. The involvement of *Dicer* in miRNA processing demonstrates a relationship with the phenomenon of RNA

<http://rfam.sanger.ac.uk/family?acc=RF00027>

View options

Alignment: Seed (14) Full (546)

Viewer: jalview

Formatting options

Alignment: Seed (14) Full (546)

Format: **Selex**

Order: **Selex** Alphabetical

Sequence: FASTA per case All upper case

Gaps: Gaps as . or - (mixed)

Download/view: Download View

Download options

Very large alignments can often cause problems for the formatting tool above. If you find that downloading or viewing a large alignment is problematic, you can also download a [gzip](#)-compressed, Stockholm-format file containing the [seed](#) or [full](#) alignment for this family.

<http://rfam.sanger.ac.uk/family?acc=RF00027>

Sequences
miRBase

Home Search Browse Genomics Help Download Submit miRBase

miRNA gene family: let-7 (189 sequences)

ID	Accession	Chromosome	Start	End	Strand	Fetch
aga-let-7	MI0001600	3R	10270708	10270797	-	<input type="checkbox"/>
age-mir-98	MI0002700					<input type="checkbox"/>
ame-let-7	MI0005726	Group8.27	75201	75300	+	<input type="checkbox"/>
bmo-let-7	MI0004968	nscaf3026	3368285	3368388	-	<input type="checkbox"/>
bta-let-7a-1	MI0005057	8	89743299	89743378	+	<input type="checkbox"/>

http://microrna.sanger.ac.uk/cgi-bin/sequences/mirna_summary.pl?fam=MIPF0000002

Stem-loop sequence MI0000001

Accession: MI0000001

ID: cel-let-7

Description: Caenorhabditis elegans let-7 stem-loop

Stem-loop

```

-----uaca gga u aua
cugu uccggugagguag agguuuuuuuuuu gg u
|||||
gaca aggcacuucauc uuuaacguuacag cc u
aguuucuaa --g u ugg acca
    
```

[Get sequence](#)

Comments

let-7 is found on chromosome X in *Caenorhabditis elegans* [1] and pairs to sites within the 3' untranslated region of these mRNAs and triggering the transition to late-larval and adult stages [2].

Genome context

Coordinates (WS190)
X: 14744094-14744192 [-]

Overlapping transcripts
intergenic

http://microrna.sanger.ac.uk/cgi-bin/sequences/mirna_entry.pl?acc=MI0000001

Data Collection & Analysis: CMfinder & CARNAC

View options

Alignment: Seed (14) Full (546)

Viewer: jalview

View

Formatting options

Alignment: Seed (14) Full (546)

Format: Selex

Order: Selex Stockholm FASTA MSF Alphabetical All upper case

Sequence: per case

Gaps: Gaps as "." or "-" (mixed)

Download/view: Download View

Generate

Download options

Very large alignments can often cause problems for the formatting tool above. If you find that downloading or viewing a large alignment is problematic, you can also download a gzip-compressed, Stockholm-format file containing the [seed](#) or [full](#) alignment for this family.

Alignment: Seed (14) Full (546)

CMfinder

CMfinder

Minimum length of motif: 30 > 15

Maximum length of motif: 100 < 150

Number of Candidates: 40 < 100

Expected fraction of sequences containing the motif: 0.8 0-1

Paste Sequences (*) in **FastA Format**
Please limit your dataset to 4-60 sequences with length < 500bp

Or Upload a FastA file (*):

Post process options: Merge motifs Remove Redundant motifs

CARNAC

CARNAC

Paste your RNA sequences in FASTA format [?]

or

upload a file

eliminate redundant sequences [?]

take GC content into account [?]

allow isolated stems (may be slow) [?]

Enter your E-mail address (optional):

CARNAC & CMfinder results

Enter a **name** for the sequences (optional) :


Paste your RNA sequences in FASTA format [?]

```
>D00721/306-255
CAACAGCGAAGCGCGCCAGGGAAACACACCAUGUGUGUAUAU
>M21212/157-106
CAACAGCGAAGCGGAACGGCGAAACACACCUUGUGUGUAUAU
>M14879/224-175
AAACAGAGAAGUCAACCAGAGAAACACACGUUGUGUAUAUU
>D00685/306-255
CAACAGCGAAGCGCGCCAGGGAAACACACCAUGUGUGUAUAU
>M17439/226-177
AAACAGAGAAGUCAACCAGAGAAACACACGUUGUGUAUAUU
```

Paste Sequences (*) in FastA Format
Please limit your dataset to
4~60 sequences with length < 500bp

```
>D00721/306-255
CAACAGCGAAGCGCGCCAGGGAAACACACCAUGUGUGUAUAU
>M21212/157-106
CAACAGCGAAGCGGAACGGCGAAACACACCUUGUGUGUAUAU
>M14879/224-175
AAACAGAGAAGUCAACCAGAGAAACACACGUUGUGUGUAUAUU
>D00685/306-255
CAACAGCGAAGCGCGCCAGGGAAACACACCAUGUGUGUAUAU
```

Or Upload a FastA file (*):

 **University of Washington**
Computer Science & Engineering

CMfinder 1.0: Results

Results will be sent to justin_slotman@yahoo.com. Please wait while we are processing your request...

Merge motifs ...

The results are now available for download: [Compressed archive \(zip\)](#) or [result directory](#)

Thank you for using CMfinder 1.0

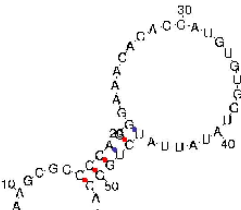
Results for job Nov270332372008

- Sequence 1 : D00721 306-255
[CT file](#) , [PS file](#) , [JPEG file](#) , [bracket notation](#) , [list of stems](#)
- Sequence 2 : M21212 157-106
[CT file](#) , [PS file](#) , [JPEG file](#) , [bracket notation](#) , [list of stems](#)
- Sequence 3 : M14879 224-175
[CT file](#) , [PS file](#) , [JPEG file](#) , [bracket notation](#) , [list of stems](#)

PS and JPEG files are generated with [NAVIEW](#).

Visualize all foldings with [RNAfamily](#)

RNAfamily is a Java applet that displays several RNA foldings at a glance. You can download the Java Virtual Machine [here](#).



Index of /CMfinder/data/ __0.85288878467

- [Parent Directory](#)
- [seq.fasta](#)
- [seq.fasta.cm.h1.1](#)
- [seq.fasta.cm.h1.2](#)
- [seq.fasta.motif.h1.1](#)
- [seq.fasta.motif.h1.2](#)
- [seq.fasta.summary](#)

Rfam and BLAT alignments

Formatting options

Alignment:	<input type="radio"/> Seed (5)	<input checked="" type="radio"/> Full (5)
Format:	FASTA	
Order:	<input checked="" type="radio"/> Tree	<input type="radio"/> Alphabetical
Sequence:	<input checked="" type="radio"/> Inserts lower case	<input type="radio"/> All upper case
Gaps:	Gaps as "-" or "." (mixed)	
Download/view:	<input type="radio"/> Download	<input checked="" type="radio"/> View

Generate

```
>D00721/306-255
CAACAGCGAAGCGCGCCAGGGAAACACACCAUGUGUGGUUAUUAUUCUGGCA
>M21212/157-106
CAACAGCGAAGCGGAAACGGCGAAACACACCUUGUGUGGUUAUUAUACCCGUUG
>M14879/224-175
AAACAGAGAAGUCAACCAGAGAAACACACGUUGUGGUUAUUAUACCGUGUA
>D00685/306-255
CAACAGCGAAGCGCGCCAGGGAAACACACCAUGUGUGGUUAUUAUUCUGGCA
>M17439/226-177
AAACAGAGAAGUCAACCAGAGAAACACACGUUGUGGUUAUUAUACCGUGUA
```

BLAT Search Genome

Genome: Assembly: Query type: Sort output: Output type:

```
>D00721/306-255
CAACAGCGAAGCGCGCCAGGGAAACACACCAUGUGUGGUUAUUAUUCUGGCA
>M21212/157-106
CAACAGCGAAGCGGAAACGGCGAAACACACCUUGUGUGGUUAUUAUACCCGUUG
>M14879/224-175
AAACAGAGAAGUCAACCAGAGAAACACACGUUGUGGUUAUUAUACCGUGUA
>D00685/306-255
CAACAGCGAAGCGCGCCAGGGAAACACACCAUGUGUGGUUAUUAUUCUGGCA
>M17439/226-177
AAACAGAGAAGUCAACCAGAGAAACACACGUUGUGGUUAUUAUACCGUGUA
```

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BLAT Search Results

ACTIONS	QUERY	SCORE	START	END	QSIZE	IDENTITY	CHRO	STRAND	START	END	SPAN
browser details	D00685/306-255	20	24	43	52	100.0%	X	-	45714471	45714490	20
browser details	D00721/306-255	20	24	43	52	100.0%	X	-	45714471	45714490	20

Conclusion

- Infernal & miRNAMiner:
 - Infernal very sensitive versus miRNAMiner
- CARNAC & CMfinder:
 - CARNAC user-friendly
 - CMfinder output difficult to interpret
- Infernal & CMfinder
 - CM-using software head to head
- BLAT and Rfam
 - Rfam full alignments include more results than corresponding BLAT searches

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